Whey Protein Determination in Milk according to the Kjeldahl method

Reference: AOAC 998.05 Non-casein Nitrogen Content of Milk; AOAC 991.21 Non-protein Nitrogen in Whole Milk; AOAC 991.20 Nitrogen (Total) in Milk; ISO 8968 - 4 FIL 20 - 4 : 2001 Non-protein Nitrogen content

Tested with VELP Scientifica DKL 20 Automatic Kjeldahl Digestion Unit (Code S30100210) and UDK 159 Automatic Kjeldahl Distillation & Titration System (Code F30200150).
Introduction

The milk proteins are the oldest and most widely consumed food proteins. There is currently a large interest in these substances, both in nutritional field and in technological application. The nitrogen compounds of milk can be divided in: casein proteins and non-casein nitrogen (NCN). Among this, there are soluble whey proteins (WP), enzymes and non-proteins nitrogen substances (NPN), composed of urea and other low molecular weight compounds, as creatine and creatinine.

Casein proteins are very important not only for the dairy products production, but also in medicine for additive and in technical use. Soluble whey proteins are non-casein proteins, defined as “not denatured milk proteins by heat treatment which can be found in serum”; they are one of the parameters for the pasteurized milk classification. In case of high-quality fresh pasteurized milk, the soluble protein serum amount must not be less than 15.5% total N.

In order to determine soluble whey proteins (WP), it’s necessary to know the amount of milk total nitrogen (N<sub>tot</sub>), non-casein nitrogen (NCN) and non-proteins nitrogen substances (NPN). WP content is calculated as difference of non-casein nitrogen (NCN) and non-proteins nitrogen (NPN) on the total nitrogen (N<sub>tot</sub>) in the milk.

Whey Protein Determination in milk according to the Kjeldahl method

Kjeldahl is nowadays the most used method for determining nitrogen and protein contents in foods and feeds, thanks to the high level of precision and reproducibility and to its simple application. The modern Kjeldahl method consists in a procedure of catalytically supported mineralization of organic material in a boiling mixture of sulphuric acid and sulphate salt at digestion temperatures higher than 400 °C. During the process the organically bonded nitrogen is converted into ammonium sulphate. Alkalizing the digested solution liberates ammonia which is quantitatively steam distilled and determined by titration.

Sample

Liquid bovine high quality milk, whole and pasteurized Protein labeled value: 3.35 g/100 ml Whey proteins content from literature: not less than 15.5%

Sample Preparation

For the determination of the non casein nitrogen (NCN) and non-proteins nitrogen (NPN) are necessary to calculate the whey proteins (WP) in the milk. The NCN and NPN are obtained separating and filtrating the milk.

Chemicals and Materials for separating and filtrating

Acetic acid solution 10% - 10 ml acetic acid diluted to 100 ml with deionized water
Sodium acetate solution 1M - 8.2 g sodium acetate diluted to 100 ml with deionized water
Trichloroacetic acid solution 20% - 20 g of TCA diluted to 100 ml with deionized water
Filter paper nitrogen free, high speed filtration

Procedure

The determination of NPN and NCN in milk involves the following steps:
- Stir the milk into a beaker using a VELP magnetic stirrer for 60 sec. at 700 rpm
- Precipitation of the casein (for NCN) or of the protein (for NPN) and filtration
- Digestion of the filtrate using DKL 20
- Distillation and titration of the sample using UDK 159
- Calculation (see the following formulas)

1. Procedure for separating NCN

Place 20 ml of milk, previously thermostated at 20 °C, in a 50 ml volumetric flask with 20 ml of deionized water. Then, put the flask at 37 °C in an Open Circulating Bath (OCB, Code F40300240) for 30 minutes. After this period, add 2 ml of acetic acid solution (10%), swirl to mix and let stand for approximately 10 minutes. Add 2 ml of sodium acetate solution 1M and let the mixture cool down to 20 °C and fill up with deionized water to the calibration mark. Then, filter through a filter paper and collect the entire filtrate.
2. Procedure for separating NPN

Place 20 ml of milk, previously thermostated at 20 °C, in a 50 ml volumetric flask and fill up with trichloracetic acid solution (20%) to the calibration mark. Let the mixture stand for 30 min. Filter with a filter paper and collect the filtrate.

Sample Digestion

Put 20 ml of filtrate into a 250 ml test tube (Code A00000144), by using a pipette. In each the test tube add:

- 2 catalyst tablets VCM (code A00000274; 3.5 g K$_2$SO$_4$, 0.1 g CuSO$_4$ 5H$_2$O Missouri)
- 4 antifoam tablets VS (Code A00000283)
- 15 ml concentrated sulphuric acid (96-98%)
- 5 ml of hydrogen peroxide (~ 30%)

Prepare some blanks with all chemicals and without sample.
Connect the Digestion Unit to a proper Aspiration Pump (JP code F30620198) and a Fume Neutralization System (SMS Scrubber code F307C0199) to neutralize the acid fumes created during digestion phase.
Digest the samples, setting the following ramps in “Customizable Methods”: for 15 minutes at 150 °C plus 45 minutes at 200 °C plus 15 minutes at 300 °C plus 60 minutes at 420 °C.

Distillation and Titration

Let the test tubes cool down to 50-60 °C.
Condition the UDK 159 unit by performing the Automatic Check up in Menu-System and a Wash down.
Distill the samples selecting the predefined method n° 1:

- H$_2$O (dilution water): 50 ml
- H$_2$SO$_4$ (0.1 N) as titrant solution
- NaOH (32%): 70 ml
- H$_3$BO$_3$ (4% with indicators): 30 ml
- Protein factor: 6.38

In UDK 159 settings, set as unit of measure mgN and %N for the final result and as sample quantity "ml".
Distillation & Titration analysis time: from 4 minutes for one test.

Typical Results

The results of the non-casein nitrogen (NCN) and non-proteins nitrogen (NPN) are calculated as percentage of nitrogen, using as sample quantity the volume of filtrate ($V_{filtrate}$) multiplied for 0.4.

1. NCN% determination in Milk

\[
NCN\% = \frac{mg\ N}{(G_{milk}/V_{sol}) \times 1000 \times V_{filtrate}} \times 100
\]

$G_{milk} =$ milk weight (20 g)
$V_{sol} =$ milk solution containing all the chemicals necessary for separating, filled up to volume (50 ml)
$V_{filtrate} =$ filtrate used to perform 1 analysis (ml)

The obtained results have been exported and multiplied for the whole milk correction factor 0.994 (for semi skimmed milk, the correction factor is 0.998).

<table>
<thead>
<tr>
<th>Filtrate quantity (ml)</th>
<th>NCN %*</th>
</tr>
</thead>
<tbody>
<tr>
<td>20.000</td>
<td>0.108</td>
</tr>
<tr>
<td>20.000</td>
<td>0.109</td>
</tr>
<tr>
<td>20.000</td>
<td>0.110</td>
</tr>
</tbody>
</table>

** Average ± SD%**

| Average ± SD% | 0.109 ± 0.001 |

| RSD% ** | 0.923 |

* already corrected with 0.994 factor  ** RSD% = (Standard Deviation x 100) / Average
2. NPN% Determination in Milk

\[ \text{NPN\%} = \frac{\text{mg N}}{[\text{G milk/V sol} \times 1000 \times V \text{ filtrate}]} \times 100 \]

- **G milk** = milk weight (20 g)
- **V sol** = milk solution containing all the chemicals necessary for separating, filled up to volume (50 ml)
- **V filtrate** = filtrate used to perform 1 analysis (ml)

<table>
<thead>
<tr>
<th>Filtrate quantity (ml)</th>
<th>NPN %*</th>
</tr>
</thead>
<tbody>
<tr>
<td>20.000</td>
<td>0.0255</td>
</tr>
<tr>
<td>20.000</td>
<td>0.0245</td>
</tr>
<tr>
<td>20.000</td>
<td>0.0258</td>
</tr>
</tbody>
</table>

Average ± SD% 0.0256 ± 0.0002

RSD% * 0.810

* RSD\% = (Standard Deviation x 100) / Average

3. Whey Proteins % Determination in Milk

\[ \text{WP \%} = \frac{\text{NCN} - \text{NPN}}{\text{N}_{\text{tot}}} \times 100 \]

<table>
<thead>
<tr>
<th>N_{\text{tot}} %</th>
<th>NCN %</th>
<th>NPN %</th>
<th>Whey Proteins %</th>
</tr>
</thead>
<tbody>
<tr>
<td>0.525</td>
<td>0.1090</td>
<td>0.0256</td>
<td>15.8</td>
</tr>
</tbody>
</table>

The complete procedure was verified by using 5 ml of glycine standard solution (3%) containing 28 mg of nitrogen, as reference substance.

The obtained recovery falls into the expected range: between 98% and 102%.

**Conclusion**

The obtained results are reliable and reproducible in accordance with the expected values, with a low relative standard deviation (RSD < 1%), that means high repeatability of the results.

Benefits of Kjeldahl method by using DKL 20 and UDK 159 are:

- High level of precision and reproducibility
- High productivity
- Worldwide official method
- Reliable and easy method
- Time saving
- Affordable equipment cost
- Moderate running costs